Rabbit Model for Germline Transmission

Rabbits injected with AAV-F.IX16 vector at doses ranging from 1 x 10¹¹ vg/kg–1 x 10¹³ vg/kg

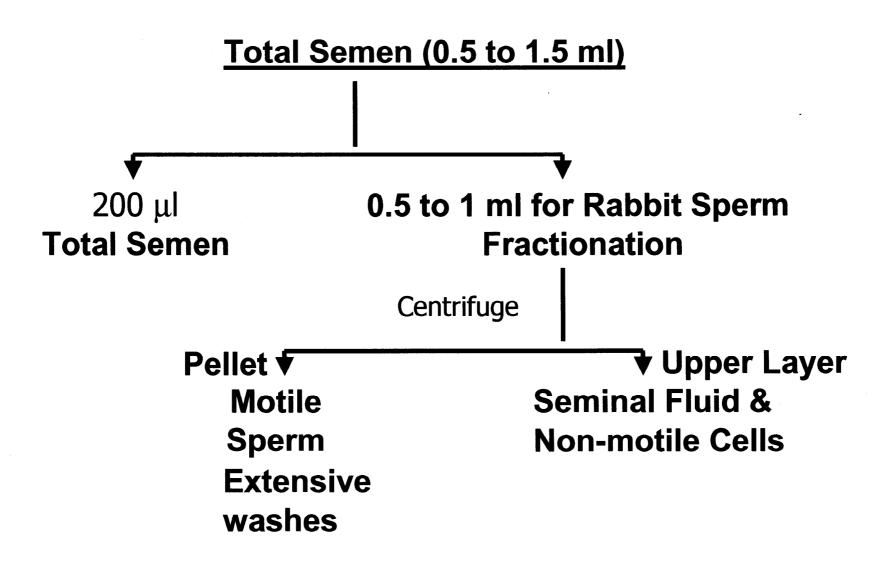
Semen collected at series of time points after injection, fractionated, DNA from total and fractions analyzed by PCR assay used to analyze human specimens

Semen Collection

Provides uncontaminated sample Artificial vagina Natural method

Disadvantages
Requires training of the male
Oestrous female

Semen Fractionation Method



PCR for Detection of Vector DNA Extraction from Semen

Southern blot of PCR product to confirm faint signal

[Probe: 0.7kb Hind III fragment of rAAV from position 905 to 1631

PCR product: position 790 to 1436 (647 bp)]

Groups of Injected Animals

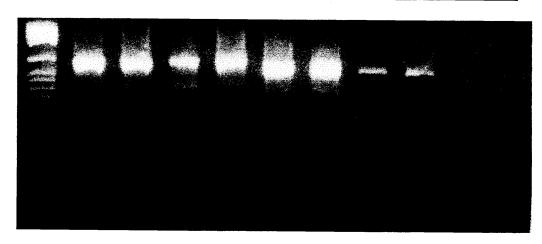
```
First cohort
  n= 12 rabbits
  5 months old, sexually mature but not experienced in
  semen collection (only later time points)
Second cohort
  n= 3 rabbits
  18 months old and experienced
  semen samples collected weekly
Third cohort
  n= 12 rabbits
  median age of 20 months (retired breeders)
```

Vector Sequences Detected in Serum 24hrs Post-Injection

M LD MD HD Water

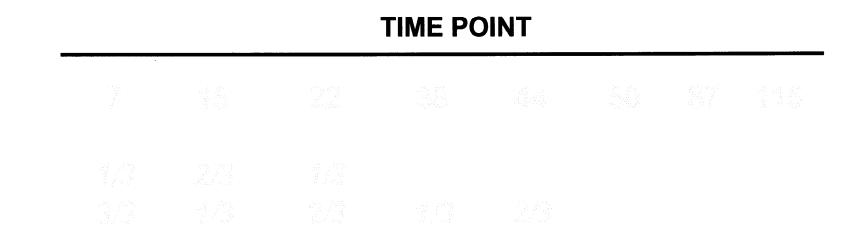


M LDIII MDIII HDIII Water



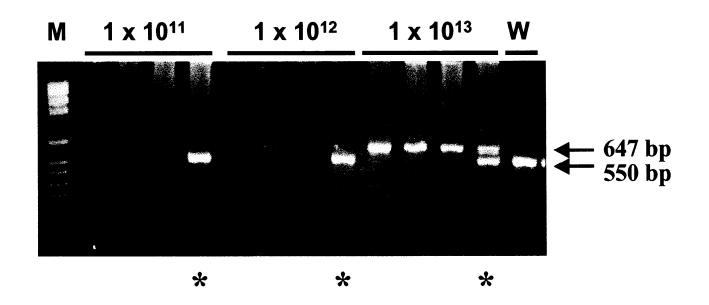
Serial Time Points

Total Semen Analysis - Experienced Rabbits



No. of Positive PCR/ No. of Total Reactions

PCR Results from DNA Extracted from Total Semen Fraction Collected 7 Days Following AAV Injection

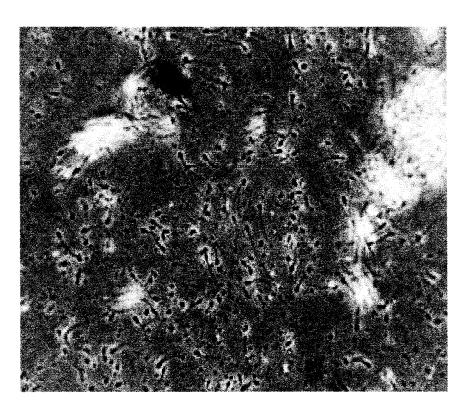


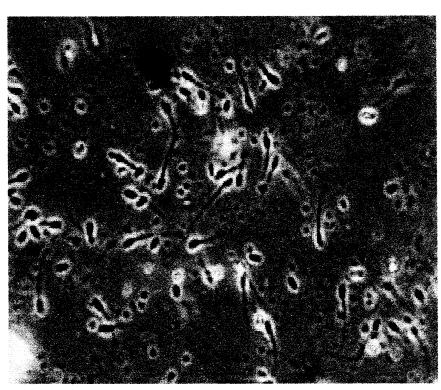
* spiked with 10 copies plasmid

Rabbit Semen Fractionation

- Optimal fractionation conditions depend on size and shape of sperm and semen pH
- Day 7 fractionation utilized parameters worked out for human semen
- Microscopic exam→contamination of "motile sperm" fraction with multiple other cell types

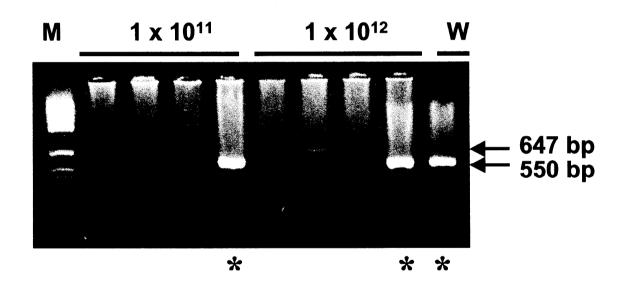
Analysis of Rabbit Semen Smear Following Fractionation





20 X 40X

PCR Results from DNA Extracted from Motile Sperm Fraction Collected 7 Days Following IV Injection of AAV-hF.IX Vector in Rabbits



* spiked with 10 copies plasmid

Comparison Between Human and Rabbit Male Germ Cells

	Humans	Rabbits	
Duration of spermatogenesis (days)	74-76	48	
Daily Sperm Production x 10 ⁶ /g of testis	s 4.4	25	
рН	7.2-8	6.6-7.5	
Volume of ejaculate (ml) comments	2-6 (3.5) much cell debris	0.4-6 (1.0) debris/gels	
Density of Sperm (x 10 ⁶ / ml)	30-120	~ 250	
motile sperm (%)	60-80	60-70	
Sperm characteristics:			
Head length	6.1	8.5	
Mid-piece length	4.7	8.8	
Total length	58.4	58.0	

Rabbit Semen Fractionation

 Specific reagents for rabbit semen fractionation (Nidacon Intl AB®)

Centrifugation speed decreased from 500g to 300g

Serial Time Points

Total Semen Analysis

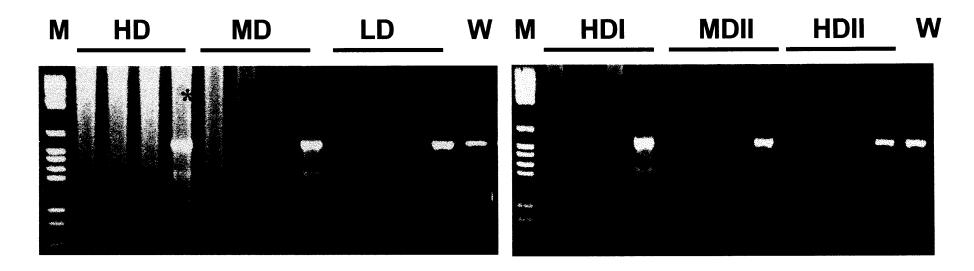
Motile Sperm Analysis

Non-motile Sperm/Seminal Fluid

NS: no sample

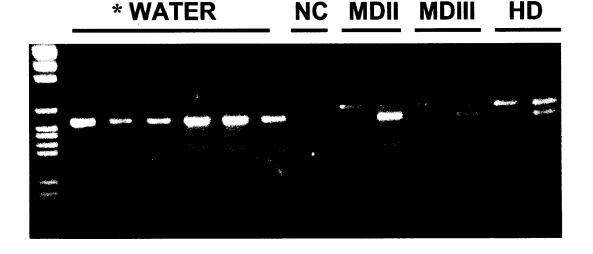
Total Semen and WBC DNA Analysis

Sample collection 3 months following injection



* spiked with 10 copies

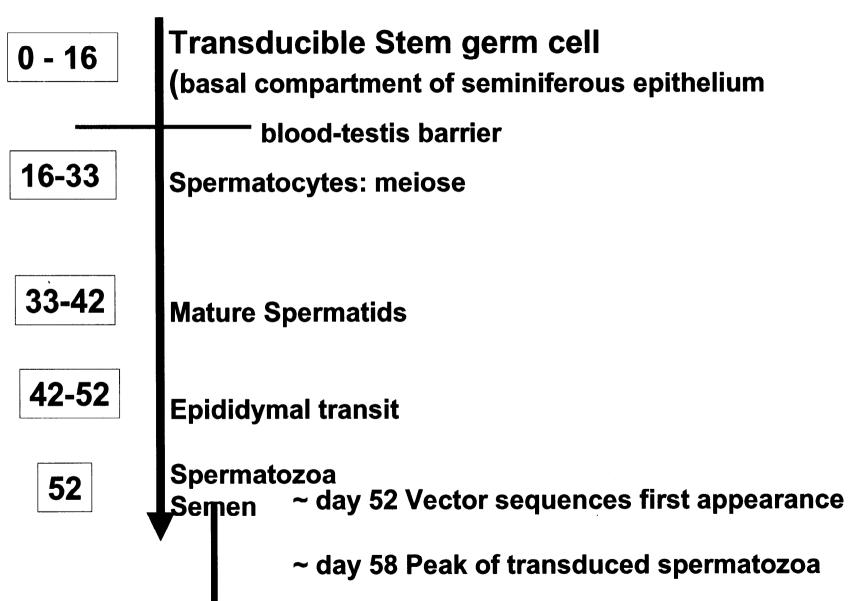
Pellet of Peripheral White Blood Cells



PCR Results from Semen Analysis-Sexually Mature but Inexperienced

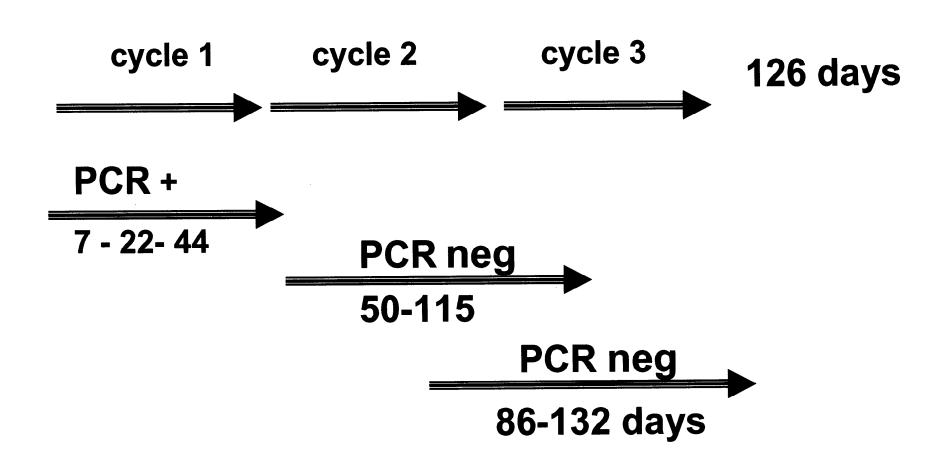


Rabbit Spermatogenesis

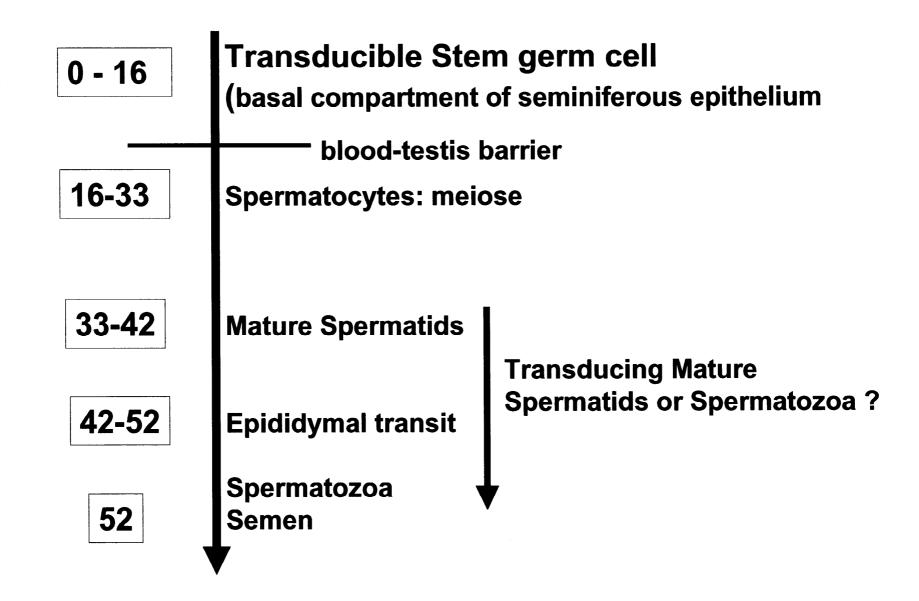


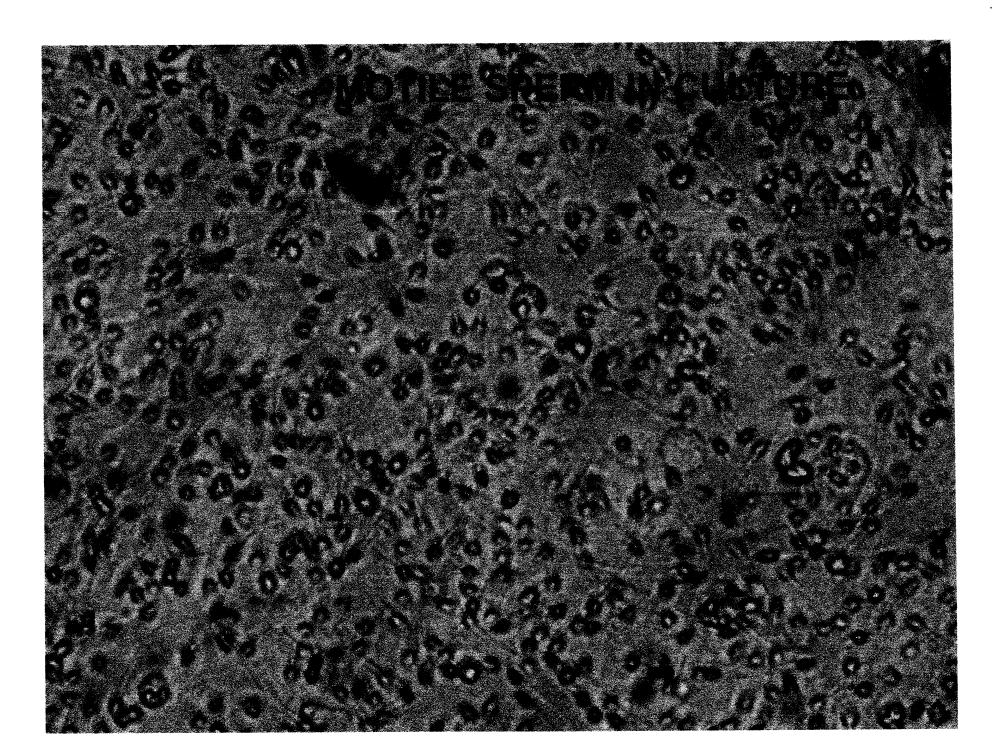
> day 70 Steady State

Rabbit Spermatogenesis



Rabbit Spermatogenesis





"In vitro" Transduction of Motile Sperm by **AAV2-CMV-GFP MOI 5,000** QuickTime™ and a Photo - JPEG decompressor QuickTime™ and a are needed to see this picture. 24 hours

Summary of Gonadal Distribution

Species	Dose	Time(Wk)	Gonads (Vector/Cell)	Semen (Vector/Cell)	Assay Sens.
Rat	1x10 ¹¹	13	negative	nd	1/15,000
	1x10 ¹²	13	negative	nd	
	1x10 ¹³	13	1/4300	nd	
Dog	3.7 – 7x10 ¹²	13	negative	negative	1/30,000
Rabbit	1x10 ¹¹	12	nd	negative	1/30,000
	1x10 ¹²	1-3	nd	1/30,000*	1/30,000
	1x10 ¹³	1-6	nd	1/30,000*	1/30,000
Monkey	7x10 ¹²	19	negative	nd	1/2,500
Human 1	2x10 ¹¹	1-10	nd	1/30,000*	1/30,000
2	2x10 ¹¹	1-12	nd	1/30,000*	1/30,000

^{*} Estimate based on qualitative assay

nd: not done

Results in assessing the risk for germline transmission of rAAV2 following <u>intramuscular</u> injection of rabbits at doses of 1 x 10¹³ vg/kg

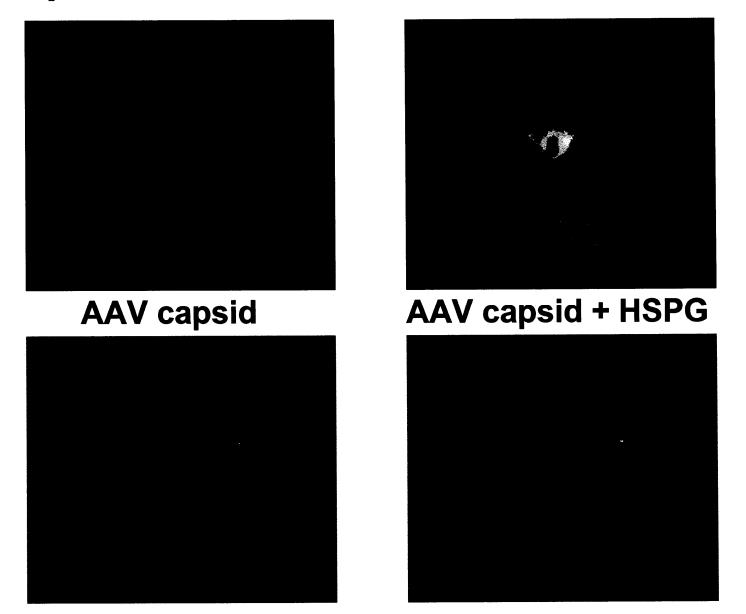
Immunofluorescence staining and Fluorescent *in situ* hybridization shows that detectable signal is localized to <u>vessel wall</u> and <u>testicular basement membrane</u>, structures rich in HSPG (known receptor for AAV2).

The detectable signal disappears with time.

No evidence of intracellular signal in testis

Mol Ther 4: 586, 2001

IF Staining for AAV Capsid and for Heparan Sulfate Proteoglycan in Testis



In Vitro Transduction of Human and Murine Cells

Human 293 cells, murine fibroblast cell line, murine spermatogonia and Sertoli cell co-cultures

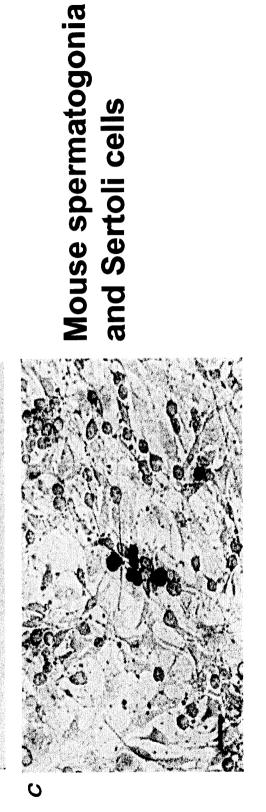
Transduced with AAV-CMV-lacZ at MOI 5000 and stain for X-gal histochemistry

Spermatogonia identified by immunostaining using Mab to germ cell nuclear antigen

Human 293 cells



Mouse fibroblast cell line (STO)



Ø

Conclusions

Intravenous administration of rAAV doses up to 1 x 10¹³ vg/kg in rabbits results in transient detectable signal in semen in a dose dependent manner

PCR positivity of the semen persists up to day 44 (high-dose animal). Follow up for ensuing ~100 days revealed no positive signal (duration of 2 times of the rabbit spermatogenesis)

Vector signal can be detected in PMBs for at least 3 months after IV injection in rabbits (in NHP up 8 months after IM injection of vector but <u>not</u> infectious after day 7)

Conclusions

Risks to partners and potential offspring can be prevented by use of barrier contraception until semen samples clear, and by sperm banking

Although it is prudent to continue studies in animal models, definitive answers to these questions can only be gained from clinical studies. Thus far there have been no other safety issues raised by hepatic artery administration of rAAV, either in animals or humans

Ongoing Experiments

Continuous follow up of kinetics of clearing, determination of anatomic localization of signal as a function of vector dose

To determine whether rAAV infectivity is detected in rabbit semen samples collected at a series of time points

To determine whether receptor for AAV-2 is present in mice, rabbit, and human spermatozoa